



Asian Journal
of
PHARMACEUTICAL RESEARCH
Journal homepage: - www.ajprjournal.com

RP-HPLC ASSAY METHOD DEVELOPMENT AND VALIDATION OF MONTELUKAST AND DOXOXYLLINE IN BULK AND PHARMACEUTICAL DOSAGE FORMS

Shajan A^{*1} and Narayanan N²

¹Research Scholar, Karpagam University, Coimbatore, Tamilnadu, India- 641 021.

²Jaya College of Pharmacy, Thiruninravur, Chennai, Tamilnadu, India- 602 024.

ABSTRACT

A simple reverse phase high performance liquid chromatographic assay method was developed and validated for the simultaneous estimation of montelukast and doxofylline from bulk and pharmaceutical dosage forms. The chromatographic separation was achieved on phenomenex C18 (250 mm × 4.6 mm id, 5 μm particle size) column by using the mobile phase composition of acetonitrile: methanol: ammonium acetate buffer (70:10:20 v/v, pH 5.5), the detection of analyte was done at 274 and 347 nm for doxofylline and montelukast respectively by PDA detector. Montelukast and doxofylline obeys Beer Lambert's law in the concentration range from 3-9 μg/ml and 120-360 μg/ml. LOD was found to be 0.0025 μg/ml and 0.0078 μg/ml and LOQ was found to be 0.0076 μg/ml and 0.0235 μg/ml for montelukast and doxofylline respectively.

Key words: RP-HPLC, Doxofylline, Montelukast, validation.

INTRODUCTION

Montelukast is a potent, selective and orally active antagonist of the cysteinyl, CysTL1, leukotriene receptor used for the treatment of asthma in children and adults [1- 3]. It is a leukotriene modifier and has clearly demonstrated the ability to ameliorate bronchoconstriction and indices of airway edema and abnormal mucus production as observed in clinical trials [4].

Doxofylline (DX) is chemically 7-(1,3-dioxolan-2-ylmethyl)-3,7-dihydro-1,3-dimethyl-1H-purine-2,6-dione. It is used as a bronchodilator in asthma and chronic obstructive pulmonary disease [5,6]. Few methods have been reported in the literature for the estimation of montelukast [7-9] and doxofylline [10,11] individually and in combined dosage forms [12]. The objective of present work is to develop and validate [13] a simple, sensitive, rapid, accurate and precise RP-HPLC assay method that can be used for the routine analysis of the formulations containing these drugs.

MATERIALS AND METHODS

Reagents and chemicals

Doxofylline and montelukast were obtained as gift samples from Burgeon Pharmaceuticals Pvt. Ltd., Chennai. Methanol, acetonitrile used as a solvent was purchased from Merck (Mumbai, India). All other reagents and solvents used were of analytical grade.

Instruments

Shimadzu HPLC instrument equipped with a PDA detector, assisted with LC- Solution software

Preparation of buffer solution

3.85 g of ammonium acetate was taken in a 1000 ml volumetric flask, 1 ml of triethyl amine was added and volume was made up to the mark with water. The pH of this buffer solution was adjusted to 5.5 with glacial acetic acid.

Preparation of mobile phase

Mobile phase was prepared having composition of acetonitrile: methanol: ammonium acetate buffer (70:10:20 v/v, pH 5.5).

Preparation of montelukast standard solution

Accurately weighed quantity of 10 mg montelukast was transferred to a 100 ml volumetric flask, dissolved in 25 ml of mobile phase and the solution was made up to the mark with mobile phase (Stock solution). From the above stock solution, 3 ml was transferred to 50 ml volumetric flask and the volume was made up with mobile phase. The solution was filtered with 0.45 μ filter and sonicated for 15 min.

Preparation of doxofylline standard solution

Accurately weighed quantity of 400 mg doxofylline was transferred to a 100 ml volumetric flask, dissolved in 25 ml of mobile phase and the solution was made up to the mark with mobile phase (Stock solution). From the above stock solution, 3 ml was transferred to 50 ml volumetric flask and the volume was made up with mobile phase. The solution was filtered with 0.45 μ filter and sonicated for 15 min.

Preparation of sample solution

Bilayer tablets of doxofylline and montelukast were prepared as per the formula developed in our previous study [14]. 20 tablets were weighed and powdered and 902.52 mg of sample was transferred into 100 ml volumetric flask and mobile phase was added to dissolve the sample. The volume was made up to the mark using mobile phase and filtered with 0.45 μ filter paper. From the above stock solution 5 ml was transferred to 100 ml volumetric flask and the volume was made up with mobile phase. The solution was filtered with 0.45 μ filter and sonicated for 15 min.

Optimized chromatographic conditions

Stationary Phase : Inertsil C18 (4.6 x 250 mm, 5 μ m)
Injector : Rheodyne

Flow rate : 1.5 ml/min
Operating temperature : Ambient temperature
Selected wave length : 274 and 347 nm for Doxofylline and montelukast respectively
Mobile phase ratio : Acetonitrile: methanol: ammonium acetate buffer (70:10:20 v/v, pH 5.5).
Injection Volume : 20 μ l
Run Time : 10 min

Validation

Linearity

The standard solution of montelukast and doxofylline was diluted with mobile phase to get concentration of 3-9 μ g/ml and 120 – 360 μ g/ml respectively. Then this solution was injected in HPLC and peak area was calculated, the calibration graph was plotted concentration versus peak area.

Accuracy

The accuracy of the method was determined by recovery experiments. The recovery studies were carried out at 50%, 100% and 150% levels by standard addition method. Standard deviation and percentage recovery was calculated.

Precision

The precision of the method was demonstrated by interday and intraday variations studies. In this study repeated injections were made at 0 h, 8 h and 16 h on the same day and different days (3 days).

Robustness

Small deliberate changes were made in the method parameters and the peak area of sample solution was calculated.

Figure 1. Calibration graph for montelukast

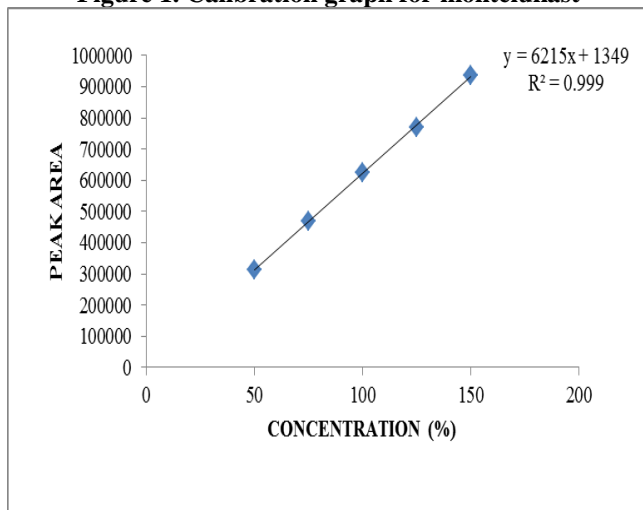


Figure 2. Calibration graph for doxofylline

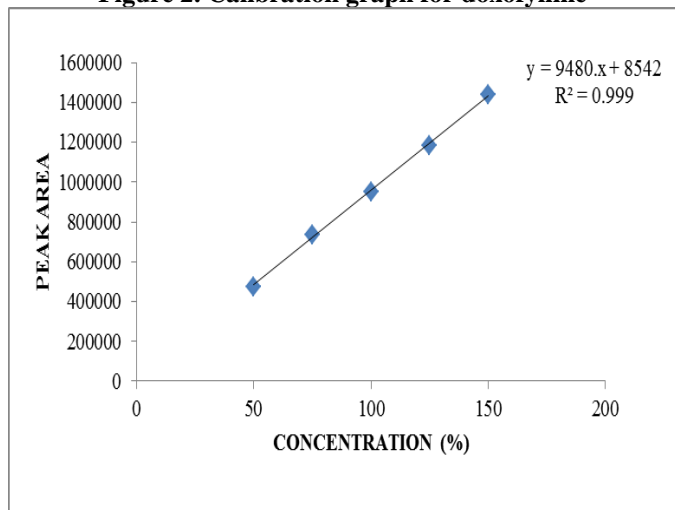


Table 1. Validation results for Montelukast and Doxofylline

Parameters	Montelukast	Doxofylline	
Specificity	No interference		
Linearity ($\mu\text{g/ml}$)	3-9	120-360	
	$R^2 - 0.999$	$R^2 - 0.999$	
Accuracy (%)	99.37	100.37	
Precision – Repeatability*(%)			
0 h	99.83 ± 0.35	99.37 ± 0.24	
8 h	99.36 ± 0.06	99.24 ± 0.12	
16 h	98.7 ± 0.01	98.85 ± 0.03	
Precision – Intermediate*(%)			
Day -1	100.07 ± 0.99	98.90 ± 0.57	
Day - 2	99.63 ± 0.03	99.22 ± 0.20	
Day - 3	98.67 ± 0.02	98.77 ± 0.02	
Instrument - 1	99.60 ± 0.15	99.02 ± 0.08	
Instrument - 2	99.86 ± 0.58	98.98 ± 0.30	
Analyst -1	99.60 ± 0.07	99.57 ± 0.01	
Analyst -2	99.96 ± 0.27	99.60 ± 0.02	
Column -1	99.61 ± 0.05	99.12 ± 0.19	
Column -2	99.51 ± 0.08	99.13 ± 0.06	
Robustness*(%)			
Flow rate	+ 10 %	99.57 ± 0.06	99.15 ± 0.09
	- 10 %	99.59 ± 0.08	99.24 ± 0.12
Mobile phase	+ 2 %	99.54 ± 0.03	98.85 ± 0.68
	- 2 %	99.57 ± 0.09	99.25 ± 0.01
Wavelength	+ 2 nm	98.02 ± 0.76	99.78 ± 0.10
	- 2 nm	97.94 ± 0.13	98.79 ± 0.16
Temperature	+ 2°C	98.91 ± 0.03	99.67 ± 0.09
	- 2°C	99.21 ± 0.03	99.81 ± 0.04
Assay	99.57 ± 0.04	99.33 ± 0.47	

*Values are expressed as mean \pm SD, n = 6

RESULTS AND DISCUSSION

The method developed in the present study using Inertsil C18 (4.6 x 250 mm, 5 µm) as stationary phase and acetonitrile: methanol: ammonium acetate buffer (70:10:20 v/v, pH 5.5) as mobile phase gave good separation of montelukast and doxofylline at 3.314 and 6.14 min respectively. The detector response of doxofylline and montelukast was found to be linear in the range of 3-9 µg/ml and 120-360 µg/ml respectively. The R² values were found to be 0.999 for doxofylline and 0.999 for montelukast (Figure 1 and 2), which indicates good linearity between concentration and peak area. The interday and intraday assay variance indicated the precision and reproducibility of the proposed method. The mean % recovery was close to 100 which indicate the

accuracy of the method. LOD was found to be 0.0025 µg/ml and 0.0078 µg/ml and LOQ was found to be 0.0076 µg/ml and 0.0235 µg/ml for montelukast and doxofylline respectively. The results are presented in table 1.

CONCLUSION

The proposed RP-HPLC method was found to be sensitive, accurate and reproducible for the quantification of doxofylline and montelukast in both bulk and solid dosage form. The excipients present in the dosage form did not interfere with the analysis. Hence the developed method can be useful for the routine quality control analysis and quantitative determinations of doxofylline and montelukast.

REFERENCES

1. Riccioni G, Vecchia RD, D'Orazio N, Sensi S and Guagnano MT. Comparison of montelukast and budesonide on bronchial reactivity in subjects with mild-moderate persistent asthma. *Pulm Pharmacol Ther*, 16, 2003, 111- 4.
2. Simons FE, Villa JR, Lee BW, Teper AM, Lyttle B, Ristizabal G, Laessing W, Schuster A, Perez-Frias J, Sekerel BE, Menten J and Leff JA. Montelukast added to budesonide in children with persistent asthma: a randomized, double blinded, crossover study. *J Pediatr.*, 138, 2001, 694-8.
3. Claesson HE and Dahlén SE. Asthma and leukotrienes: ntileukotrienes as novel anti-asthmatic drugs. *J Int Med*, 245, 1999, 205-27.
4. Valacer DJ. New treatments for asthma:the role of leukotriene modifier agents. *J Natl Med Assoc*, 91, 1999, 26S- 39S.
5. Budavari S, Eds., In:, 3rd Ed. Merck and Copress., Inc., Whitehouse Station, The Merck Index, NJ. 2001, 86.
6. Reynolds JEF and Martindale. The Extra Pharmacopoeia 29th ed. The Pharmaceutical press. London. 1989,1492.
7. Smita Patil, Pore YV, Kuchekar BS, Aruna Mane and Khire UG. Determination of Montelukast sodium and Bambuterol hydrochloride in tablets using RP-HPLC. *Indian J Pharm Sci*, 71(1), 2009.
8. Rashmitha, Joseph ST, Srinivas CH, Srinivas N, Ray UK, Hemant KS and Mukkanti K. A validated RP- HPLC method for the Determination of impurities in montelukast sodium. *E-J Chemistry*, 7(2), 2010, 555-563.
9. Sing RM, Saini PK, Mathur SC, Sing GV, and Lal B. Development and validation of a RP-HPLC method for estimation of montelukast sodium in bulk and in tablet dosage form. *Curr Trends in Biotech Pharm*, 72(2), 2010, 235-237.
10. Nimmagada S, lakshmi narasu M and Praba Shankar B. Development and validation of a sensitive LC-MS/MS method with eletrospray ionization for quantification of Doxofylline in human serum. *Biomed Chromatography*, 22(6), 2008, 654-61.
11. Kamila MM, Mondal N and Ghosh LK. Development and validation of Spectrophotometric method for estimation of Doxofylline in bulk drug and pharmaceutical preparation. *Indian J Chem Tech*, 14, 2007, 523-5.
12. Giriraj P and Shajan A. Simultaneous Estimation and Method Validation of Montelukast Sodium and Doxofylline in Solid Dosage form by RP-HPLC. *Int J Chem Pharm Sci*, 2(1), 2011, 31-36.
13. ICH, Q2B, Validation of analytical procedures :Methodology, In proceedings of the International conference on Harmonisation. Geneva.
14. Shajan A, Narayanan N, Reema N, Aiswarya G. Design and evaluation of bilayer tablets of doxofylline HCl and montelukast sodium for the treatment of asthma. *Int J Pharm Ind Res*, 3(1), 2013, 70-76.